

## p53, Oxidative Stress, and Aging

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### Abstract

Mammalian aging is associated with elevated levels of oxidative damage of DNA, proteins, and lipids as a result of unbalanced prooxidant and antioxidant activities. Accumulating evidence indicates that oxidative stress is a major physiological inducer of aging. p53, the guardian of the genome that is important for cellular responses to oxidative stresses, might be a key coordinator of oxidative stress and aging. In response to low levels of oxidative stresses, p53 exhibits antioxidant activities to eliminate oxidative stress and ensure cell survival; in response to high levels of oxidative stresses, p53 exhibits prooxidative activities that further increase the levels of stresses, leading to cell death. p53 accomplishes these context-dependent roles by regulating the expression of a panel of genes involved in cellular responses to oxidative stresses and by modulating other pathways important for oxidative stress responses. The mechanism that switches p53 function from antioxidant to prooxidant remains unclear, but could account for the findings that increased p53 activities have been linked to both accelerated aging and increased life span in mice. Therefore, a balance of p53 antioxidant and prooxidant activities in response to oxidative stresses could be important for longevity by suppressing the accumulation of oxidative stresses and DNA damage. *Antioxid. Redox Signal.* 15, 1669–1678.

### p53 Is a Critical Tumor Suppressor

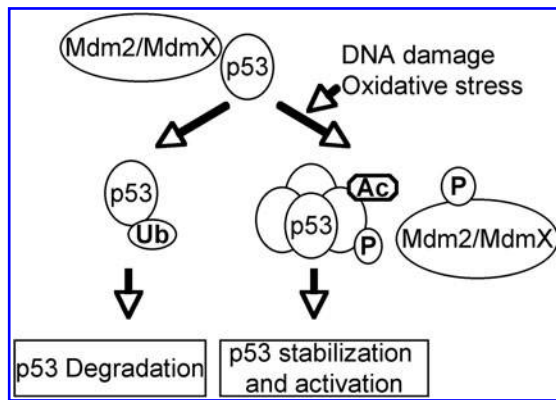
THE CRITICAL TUMOR SUPPRESSOR p53 plays important roles in cell-cycle arrest, apoptosis, senescence, or differentiation in response to various genotoxic and cellular stresses, including oxidative stress (73, 102, 133). As a guardian of the genome, the inactivation of wild-type p53 function by direct gene mutation or disruption of pathways important for p53 activation is a prerequisite for the development of most human cancers (35, 92, 127). As a transcription factor, p53 consists of two N-terminal transactivation domains, a core DNA-binding domain and a C-terminal oligomerization domain (55, 92). Because of its potent activity in inducing apoptosis and senescence, the p53 stability and activity are tightly regulated by posttranslational mechanisms (47, 51, 129). In the absence of stresses, p53 is inactive and unstable because of its interaction with Mdm2 and MdmX, which inactivate p53 and ubiquitinate p53 for proteasome-dependent degradation (Fig. 1). In response to stresses, p53 is modified posttranslationally through phosphorylation, acetylation, methylation, and sumoylation at various sites, disrupting the interaction between p53 and its negative regulators, leading to the activation and stabilization of p53 (68, 85, 104).

As a transcription factor, p53 can directly regulate the expression of hundreds of genes, products of which mediate various p53-dependent functions (Fig. 2) (43, 53, 69). For example, p21 and 14-3-3 $\sigma$  are responsible for p53-dependent

cell-cycle arrest (30, 31, 50); p53 can also induce embryonic stem (ES) cell differentiation by suppressing the expression of Nanog, which is required for the self-renewal of ES cells (64). In response to high levels of DNA damage, p53 induces apoptosis and senescence by upregulating apoptotic genes such as *Noxa* and *Puma* (66, 71). These functions of p53 prevent the passage of DNA damage to the daughter cells and thus maintain genomic stability. In response to oxidative stresses, p53 activates the transcription of a number of genes involved in regulating oxidative stress, such as Sestrin, glutathione peroxidase (GPX), aldehyde dehydrogenase (ALDH), and tumor protein 53-induced nuclear protein 1 (TP53INP1) (14, 16, 115, 130). p53 can also regulate the cellular oxidative stress levels by modulating glycolysis through inducing the expression of TIGAR (TP53-induced glycolysis and apoptosis regulator) and suppressing the expression of phosphoglycerate mutase (PGM) (9, 58).

### p53 and Aging

Recent studies have functionally linked p53 to aging in various organisms (Fig. 3). The p53 orthologue in *Caenorhabditis elegans*, Cep-1, is involved in negatively regulating the life span of the worm, because the reduced expression of Cep-1 results in increased longevity (4). Expression of dominant-negative versions of *Drosophila melanogaster* p53 (Dmp53) in adult neurons extends the life span and increases the genotoxic stress



**FIG. 1. Activation of p53 in response to DNA damage and oxidative stresses.** In the absence of stresses, the negative regulators of p53, such as Mdm2/MdmX, suppress p53 activity and induce its degradation. In response to DNA damage and oxidative stress, p53 and its negative regulators are posttranslationally modified, leading to p53 activation by disrupting the interaction between p53 and its negative regulators.

resistance in the fly (8). Because the expression of the dominant-negative Dmp53 does not further increase the life span of flies that are calorie restricted, these findings suggest that p53 is involved in mediating the calorie-restricted life span in flies. However, mutagenesis studies in *C. elegans* show that certain mutations extending the life span increase activities of p53 and cancer resistance (94). Therefore, increased p53 activities are associated with both accelerated aging and increased life span in *C. elegans*.

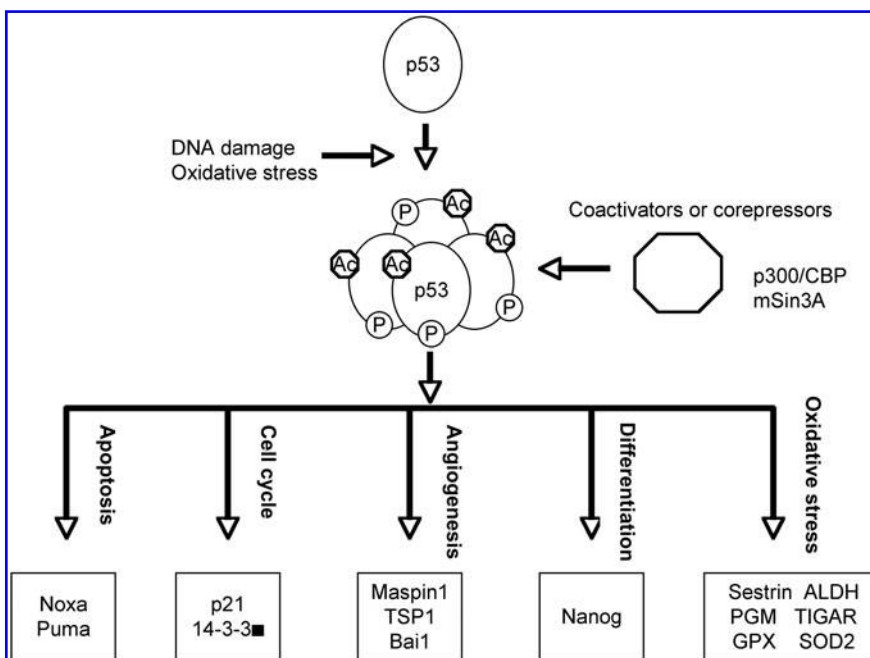
A similarly complicated scenario is also observed when studying the roles of p53 in mammalian aging. One mouse model, in which the N-terminus of p53 is truncated, exhibits increased p53 activities and accelerated aging (119). However, because of the large deletion of the genomic DNA upstream of

p53 that contains 24 genes (40), it remains unclear whether any of these deleted genes is responsible for these aging phenotypes. The potential involvement of N-terminus-truncated p53 in aging is further supported by the overexpression of the N-terminus-deleted p53 isoform p44 in mice, leading to accelerated aging (72). This study suggests that p44 modulates the life span by inhibiting the PTEN and IGF signal pathways (39, 75, 110). To link p53 to aging in humans, a recent study shows that polymorphism of p53 at codon 72 (arginine-to-proline substitution) reduces p53 activities, correlating with increased life span but also with higher cancer risk in older individuals (120). Therefore, it has been suggested that p53 might suppress cancer at the cost of longevity.

The notion that increased p53 activity induces aging in mice is challenged by recent studies of mouse models with increased p53 activities. For example, mice with a hypomorphic mutation in Mdm2 exhibit increased p53 activity but normal life span (78). In addition, mice with an additional copy of p53 and ARF exhibit an enhanced expression of antioxidant activity and decreased levels of endogenous oxidative stresses, correlating with increased life span (74). Therefore, the increased antioxidant activity of p53 in these transgenic mice prevents the accumulation of oxidative stresses to the high levels required to induce p53-dependent apoptosis and senescence, thus delaying aging in these mice. In summary, the functions of p53 in aging are complex and could be context dependent. In this context, mild and transient activation of p53 in response to a low dosage of oxidative stress could protect cells from oxidative damage. In contrast, persistent activation of p53 in response to high levels of oxidative stresses can result in cell death and organismal aging. In further support of this notion, persistent activation of p53 depletes adult stem cells primarily through p53-dependent apoptosis (64).

### Oxidative Stress and Aging

The free radical hypothesis remains the most well-established theory on the mechanism of aging (46). The increased



**FIG. 2. p53 target genes are mediators of various p53-dependent functions in response to DNA damage and oxidative stresses.**

Organism	Genetic modification	Impact on p53 activity	Impact on lifespan
<i>C. elegans</i>	Reduced expression	Reduced	Extended
<i>Drosophila</i>	Dominant negative	Reduced	Extended
Mouse	N-terminus deleted	Increased	Shortened
	p44	Increased	Shortened
	p53 <sup>T21S23D</sup> knock-in	Increased	Shortened
	Transgenic Arf or p53	Increased	No impact
	Transgenic Arf and p53	Increased	Extended
	Hypomorphic Mdm2	Increased	No impact

FIG. 3. Summary of the modulation of p53 effects on the lifespan of various organisms.

ROS production and a decreased antioxidant capacity are thought to contribute to the aging process by oxidative modification of different macromolecules, such as lipids, proteins, and genomic DNA (12, 20, 25, 62, 63, 65, 96, 109, 117). In the context of DNA, oxidative damage to mitochondrial and nuclear DNA is significantly increased in different tissues in old rats and mice (20, 45, 61, 67, 76, 82, 116). Levels of lipid-peroxidation products are also increased with aging (44, 83, 87, 97, 108, 113, 119, 123). In addition, aging-related oxidative modification of different proteins causes changes in protein structure, enzyme activities, transcriptional activities, and signal-transduction pathways (32, 70, 103, 111, 112, 124), leading to age-related diseases. In summary, the levels of oxidative damage are increased during aging in various organisms, including *C. elegans* (11, 52, 121), flies (3, 64), and mice (22, 74, 79).

Free radicals are physiologic byproducts of metabolism and are rapidly eliminated by various antioxidant enzymes in cells (23). For example, the antioxidant enzymes, including superoxide dismutase (SOD), catalase, and peroxiredoxins, convert superoxide to hydrogen peroxide and eventually to water (5, 19, 99). SODs catalyze the breakdown of the superoxide anion into oxygen and hydrogen peroxide. Mice lacking SOD2 develop neurologic defects and die soon after birth because of excessive mitochondrial production of ROS (77); mice lacking SOD1 are viable but have numerous pathologies and a reduced life span (98). Catalase converts hydrogen peroxide into water and oxygen (19, 132). Humans and mice deficient in catalase can still efficiently remove H<sub>2</sub>O<sub>2</sub>, implying that other enzymes are also involved in this reaction (72, 88). Peroxiredoxins catalyze the reduction of hydrogen peroxide, organic peroxide, and peroxynitrite (99). These enzymes can be divided into three classes: typical 2-cysteine peroxiredoxins, atypical 2-cysteine peroxiredoxins, and 1-cysteine peroxiredoxins (128). Mice lacking peroxiredoxins 1 and 2 have a shortened life span (55, 86). Together, these findings underscore the importance of antioxidant enzymes in preventing aging processes. In further support of this notion, a diet rich in the building-block nutrients of antioxidant enzymes, including cofactors for SOD (manganese, zinc, and copper), show beneficial effects on delaying aging (1, 24, 49, 59, 81, 106).

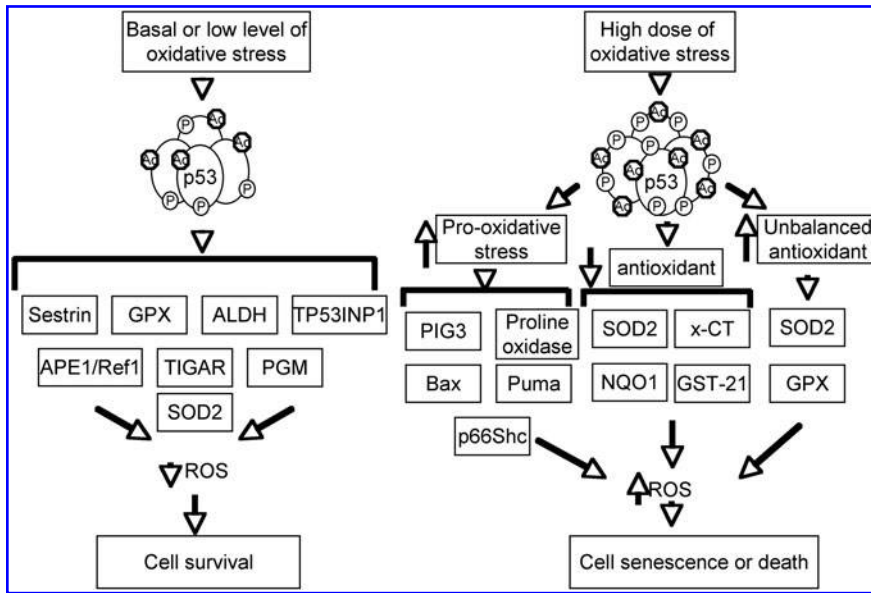
In further support of the notion that oxidative stress is an inducer of aging, treatment with antioxidants can increase the life spans of various organisms and has a beneficial impact on aging-related diseases (6, 29, 38, 57, 114, 119). A low dose of dietary supplement with antioxidants partially mimics the effects of caloric restriction and delays aging in mice (6), and long-term treatment with free radical scavenging Schisandrin B, a dibenzocyclooctadiene derivative isolated from the fruit of *Schisandra chinensis*, delays aging-related functional impairment in various organs and improves the survival rate of aging mice (114). A dietary supplement of cysteine, which is required for the synthesis of the primary antioxidant glutathione, has clear benefits in delaying some aspects of aging (29). However, clinical trials have also found no significant beneficial effects of supplementation with antioxidant vitamin E, indicating that not all antioxidants have antiaging activities (55, 107, 125).

### p53 and Oxidative Stress

ROS levels have a significant impact on cell growth, survival and development, and tumorigenesis (17). p53 plays key and complex roles in cellular responses to oxidative stresses (84, 100). In response to low levels of oxidative stresses, p53 plays primarily antioxidant roles. In this context, a number of p53 target genes, including Sestrin, glutathione peroxidase (GPX), and aldehyde dehydrogenase (ALDH), are involved in reducing oxidative stresses (Fig. 4). For example, *Sestrin* protects the cells from hydrogen peroxide-induced damage by generation of peroxiredoxins (14). GPX is a primary antioxidant enzyme that scavenges hydrogen peroxide or organic hydroperoxides (115). Aldehyde dehydrogenase (ALDH) also contributes to the antioxidant function of p53 (130).

p53 can also reduce the intracellular levels of ROS by regulating cellular metabolism. In this context, p53 induces the expression of TIGAR (TP53-induced glycolysis and apoptosis regulator), which slows glycolysis and promotes the production of NADPH to decrease ROS levels (9). In addition, p53 suppresses the expression of phosphoglycerate mutase (PGM), leading to a decrease of pyruvate required for oxidative respiration in mitochondria and thus reduced ROS production (10, 74).

In response to high levels of oxidative stress, p53 exhibits prooxidative activities by turning on prooxidative genes such as *PIG3* and *proline oxidase* (27, 95). Overexpression of these genes leads to higher levels of oxidative stress. In addition, p53 induces the expression of BAX and PUMA, which induce apoptosis through the release of cytochrome *c* from mitochondria (66, 71). The prooxidative activities of p53 also include the inhibition of the expression of antioxidant genes, leading to increased cellular oxidative stresses to induce apoptosis. For example, p53 could repress the expression of SOD2 and Nrf2, resulting in sensitivity to oxidative stress or inducing apoptosis (28, 34, 91). Interestingly, p53-induced upregulation of MnSOD and GPX, but not catalase, increases oxidative stress and apoptosis (54), suggesting that the balance of antioxidant enzyme and oxidative stress is important for cell survival. In summary, p53 plays important but context-dependent roles in regulating cellular oxidative stresses, and the levels of oxidative-stress damage dictate whether the p53 behavior is that of a protector or a killer (100).



**FIG. 4. Context-dependent roles of p53 in cellular responses to oxidative stresses by turning on distinct target genes.** At basal or low levels of oxidative stress, p53 regulates the expression of Sestrin, GPX, ALDH, TP53INP1, SOD2, TIGAR, and PGM to eliminate ROS, and therefore, promotes cellular survival. In response to high levels of oxidative stress, p53 induces the expression of prooxidative genes and suppresses the expression of antioxidant genes to increase ROS levels and promote apoptosis. Unbalanced antioxidants can also induce ROS to promote cell death.

### p53 Interacts with Other Pathways Involved in Oxidative Stress and Aging

In addition to its direct regulation of genes involved in oxidative stresses, p53 also interacts with other pathways that are involved in aging and oxidative stresses, which are summarized here (Fig. 5).

#### Sirt1

The *Sirt1* gene encodes the NAD-dependent histone deacetylase, which is important for the longevity in yeast and mammalian species by calorie restriction (42, 60, 64, 122). *Sirt1* can deacetylate and inactivate p53, leading to impaired cell growth arrest and apoptosis in response to oxidative stresses (101). In addition, the expression of a dominant-negative version of Sirt1 increases the cellular sensitivity to oxidative stress, further indicating its antioxidant roles in cellular responses to oxidative stresses. However, the roles of *Sirt1* in suppressing p53 in response to oxidative stresses remain to be fully established. In contrast to the prediction that Sirt1 deficiency would increase p53 activity, recent studies indicate that deficiency of Sirt1 extends the replicative capacity of mouse embryonic fibroblasts (MEFs) under the conditions of chronic oxidative stress due to the inefficient activation of p53 (21). However, the physiological relevance of replicative senescence in aging is not clear, because it primarily reflects a cell-culture phenomenon in the presence of nonphysiologically high levels of oxygen. Because Sirt1 is an NAD-dependent deacetylase, and NAD levels are regulated by cellular metabolism and levels of ROS, these findings implicate a complex functional interaction of p53, Sirt1, oxidative stresses, and aging.

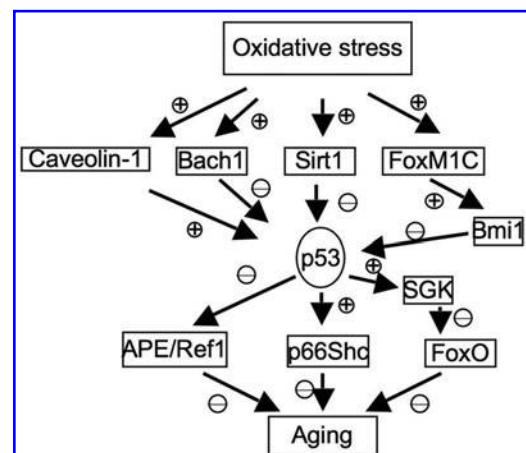
#### p66Shc

p66Shc, a downstream target of p53, is indispensable for p53-dependent elevation of intracellular oxidative stresses and apoptosis (118). p66Shc is a splice variant of p52Shc/p46Shc, a cytoplasmic signal transducer involved in the transmission of mitogenic signal from activated receptors to

Ras (93). However, p66Shc is not involved in regulating Ras signal but instead is involved in inducing apoptosis in response to oxidative stresses (80). The important role of p66Shc in oxidative stresses and aging is indicated by the findings that ablation of p66Shc enhances cellular resistance to apoptosis induced by oxidative stresses and extends the life span of p66Shc-deficient mice (79). In this context, cytochrome *c* release after oxidative signals is impaired in p66Shc-deficient cells (90). Therefore, p66Shc functionally links p53 to oxidative stress response and aging.

#### FoxO

Forkhead box O (FoxO) transcription factors are important mediators of the PI3K/Akt signaling pathway and regulate the cellular responses to oxidative stresses and the life span (56, 105). p53 negatively regulates the activities of FoxO by inducing the expression of serum- and glucocorticoid-inducible kinase (SGK), a negative regulator of FoxO and PTEN (37). In addition, Sirt1 can deacetylate FoxO3 and FoxO4, thus



**FIG. 5. Functional interaction between p53 and other pathways important for oxidative stress response and aging.**



attenuating FoxO-induced apoptosis and cell-cycle arrest (41). Therefore, the balance of the functional interaction among Sirt1, FoxO, and p53 might play important roles in regulating oxidative stresses and aging.

#### APE/Ref1

The expression of APE/Ref1 is decreased in senescent human bone marrow-derived mesenchymal stem cells (hBMSCs) with increased endogenous ROS levels. Overexpression of APE1/Ref-1 suppresses superoxide production and decreases senescence in hBMSCs (48). In addition, aging mice have an impaired induction of APE in response to oxidative damage (15). The activities of APE/Ref1 are negatively regulated by p53 (131), implicating another pathway for p53 to modulate oxidative stresses and aging.

#### Caveolin-1

Expression of Caveolin-1 is induced in fibroblasts undergoing oxidative stress-induced senescence, and the antioxidant prevents the senescence and upregulation of Caveolin-1 (36, 126). Overexpression of Caveolin-1 in MEF induces the premature senescence through a p53-p21-dependent pathway, suggesting that Caveolin-1 could activate p53-dependent premature senescence after oxidative stresses (36). In this context, Caveolin-1 binds to Mdm2 and disrupts the binding of Mdm2 to p53, leading to the activation of p53 in response to oxidative stresses. The activation of p53 and induction of premature senescence are compromised in the Caveolin-1-null MEFs, confirming that Caveolin-1 is an upstream activator of p53 in response to oxidative stresses (7).

#### FoxM1C-Bmi1 pathway

Bmi1 is a negative regulator of the *Ink4a/Arf* and p53; FoxM1C induces the expression of Bmi1 to prevent the oxidative stress-induced cellular senescence by inhibiting the expression of p53 (13, 18, 33, 89). Bmi1 is important to repress the prooxidant activities of p53 in neurons and to suppress oxidative stress-induced apoptosis and premature aging-like phenotypes (18). In addition, targeted depletion of Bmi1 sensitizes tumor cells to p53-mediated apoptosis in response to radiation therapy (2).

#### Bach1

For transcription factors, the recruitment of co-activators or co-repressors to p53 target promoters is critical for p53-dependent transcription. Bach1 is induced by oxidative stresses and forms a complex with p53, histone deacetylase 1, and nuclear co-repressor N-coR, promoting histone deacetylation and suppression of certain p53 target genes (26). In this context, Bach1 inhibits oxidative stress-induced cellular senescence by disrupting p53-dependent gene expression (26).

#### Conclusion

The accumulation of oxidative stress and oxidative damage is a major inducer of aging. Many pathways involved in cellular responses to oxidative stresses regulate the aging process and the life spans of various organisms. p53 plays important but context-dependent roles in cellular responses to low or high levels of oxidative stresses. In response to low levels of

oxidative stresses, p53 exhibits antioxidant activities and promotes cellular survival; in response to high levels of oxidative stresses, p53 exhibits prooxidative activities to induce cellular apoptosis. Both functions of p53 can prevent the accumulation of oxidative damage in cells and thus maintain genomic stability. p53 accomplishes these functions by direct transcriptional regulation of genes involved in oxidative-stress responses or modulating other pathways important in oxidative-stress responses.

Consistent with its context-specific roles in oxidative-stress responses, the roles of p53 in aging appear to be complex as well. In this context, increased p53 activities can accelerate aging in some transgenic mouse models but not in others (72, 74, 78, 119). In addition, the increase of the gene dosage of ARF and p53 does not promote aging but increases the life span of transgenic mice (74). Therefore, the roles of p53 in aging could also be context dependent. The accumulation of oxidative stresses in old mice could turn on the apoptotic or senescent roles of p53, thus promoting the aging process. However, increased dosages of p53 and ARF could ensure more efficient elimination of oxidative stress and thus prevent the accumulation of oxidative stresses to high levels in old mice. In support of this notion, a significant reduction of DNA damage occurs in old transgenic mice with additional copies of p53 and ARF (74). p53 primarily plays a protective role to increase the life span in these transgenic mice. Therefore, further elucidation of the mechanism that governs the context-dependent roles of p53 in oxidative-stress responses and the functional outcomes of the interaction between p53 and other pathways involved in cellular responses to oxidative stresses will shed light on its role in aging.

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#### References

1. Airede AK. Copper, zinc and superoxide dismutase activities in premature infants: a review. *East Afr Med J* 70: 441–444, 1993.
2. Alajez NM, Shi W, Hui AB, Yue S, Ng R, Lo KW, Bastianutto C, O'Sullivan B, Gullane P, and Liu FF. Targeted depletion of BMI1 sensitizes tumor cells to P53-mediated apoptosis in response to radiation therapy. *Cell Death Differ* 16: 1469–1479, 2009.
3. Arking R, Buck S, Berrios A, Dwyer S, and Baker GT 3rd. Elevated paraquat resistance can be used as a bioassay for longevity in a genetically based long-lived strain of *Drosophila*. *Dev Genet* 12: 362–370, 1991.
4. Arum O and Johnson TE. Reduced expression of the *Caenorhabditis elegans* p53 ortholog cep-1 results in increased longevity. *J Gerontol A Biol Sci Med Sci* 62: 951–959, 2007.
5. Bannister JV, Bannister WH, and Rotilio G. Aspects of the structure, function, and applications of superoxide dismutase. *CRC Crit Rev Biochem* 22: 111–180, 1987.
6. Barger JL, Kayo T, Vann JM, Arias EB, Wang J, Hacker TA, Wang Y, Raederstorff D, Morrow JD, Leeuwenburgh C, Allison DB, Saupé KW, Cartee GD, Weindruch R, and Prolla TA. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS One* 3: e2264, 2008.

7. Bartholomew JN, Volonte D, and Galbiati F. Caveolin-1 regulates the antagonistic pleiotropic properties of cellular senescence through a novel Mdm2/p53-mediated pathway. *Cancer Res* 69: 2878–2886, 2009.
8. Bauer JH, Poon PC, Glatt-Deeley H, Abrams JM, and Helfand SL. Neuronal expression of p53 dominant-negative proteins in adult *Drosophila melanogaster* extends life span. *Curr Biol* 15: 2063–2068, 2005.
9. Bensaad K, Tsuruta A, Selak MA, Vidal MN, Nakano K, Bartrons R, Gottlieb E, and Vousden KH. TIGAR, a p53-inducible regulator of glycolysis and apoptosis. *Cell* 126: 107–120, 2006.
10. Bensaad K and Vousden KH. p53: new roles in metabolism. *Trends Cell Biol* 17: 286–291, 2007.
11. Berdichevsky A, Viswanathan M, Horvitz HR, and Guarante L. *C. elegans* SIR-2.1 interacts with 14-3-3 proteins to activate DAF-16 and extend life span. *Cell* 125: 1165–1177, 2006.
12. Bergamini E, Bizzarri R, Cavallini G, Cerbai B, Chiellini E, Donati A, Gori Z, Manfrini A, Parentini I, Signori F, and Tamburini I. Ageing and oxidative stress: a role for dolichol in the antioxidant machinery of cell membranes? *J Alzheimers Dis* 6: 129–135, 2004.
13. Bruggeman SW, Valk-Lingbeek ME, van der Stoop PP, Jacobs JJ, Kieboom K, Tanger E, Hulsman D, Leung C, Arsenijevic Y, Marino S, and van Lohuizen M. Ink4a and Arf differentially affect cell proliferation and neural stem cell self-renewal in Bmi1-deficient mice. *Genes Dev* 19: 1438–1443, 2005.
14. Budanov AV, Sablina AA, Feinstein E, Koonin EV, and Chumakov PM. Regeneration of peroxiredoxins by p53-regulated sestrins, homologs of bacterial AhpD. *Science* 304: 596–600, 2004.
15. Cabelof DC, Raffoul JJ, Ge Y, Van Remmen H, Matherly LH, and Heydari AR. Age-related loss of the DNA repair response following exposure to oxidative stress. *J Gerontol A Biol Sci Med Sci* 61: 427–434, 2006.
16. Cano CE, Gommeaux J, Pietri S, Culcasi M, Garcia S, Seux M, Barelier S, Vasseur S, Spoto RP, Pebusque MJ, Dusetti NJ, Iovanna JL, and Carrier A. Tumor protein 53-induced nuclear protein 1 is a major mediator of p53 antioxidant function. *Cancer Res* 69: 219–226, 2009.
17. Chao C, Hergenbahn M, Kaeser MD, Wu Z, Saito S, Iggo R, Hollstein M, Appella E, and Xu Y. Cell type- and promoter-specific roles of Ser18 phosphorylation in regulating p53 responses. *J Biol Chem* 278: 41028–41033, 2003.
18. Chatoo W, Abdouh M, David J, Champagne MP, Ferreira J, Rodier F, and Bernier G. The polycomb group gene Bmi1 regulates antioxidant defenses in neurons by repressing p53 pro-oxidant activity. *J Neurosci* 29: 529–542, 2009.
19. Chelikani P, Fita I, and Loewen PC. Diversity of structures and properties among catalases. *Cell Mol Life Sci* 61: 192–208, 2004.
20. Chen JH, Hales CN, and Ozanne SE. DNA damage, cellular senescence and organismal ageing: causal or correlative? *Nucleic Acids Res* 35: 7417–7428, 2007.
21. Chua KF, Mostoslavsky R, Lombard DB, Pang WW, Saito S, Franco S, Kaushal D, Cheng HL, Fischer MR, Stokes N, Murphy MM, Appella E, and Alt FW. Mammalian SIRT1 limits replicative life span in response to chronic genotoxic stress. *Cell Metab* 2: 67–76, 2005.
22. Collins AR, Lyon CJ, Xia X, Liu JZ, Tangirala RK, Yin F, Boyadjian R, Bikineyeva A, Pratico D, Harrison DG, and Hsueh WA. Age-accelerated atherosclerosis correlates with failure to upregulate antioxidant genes. *Circ Res* 104: e42–e54, 2009.
23. Davies KJ. Oxidative stress: the paradox of aerobic life. *Biochem Soc Symp* 61: 131, 1995.
24. Davis CD and Feng Y. Dietary copper, manganese and iron affect the formation of aberrant crypts in colon of rats administered 3,2'-dimethyl-4-aminobiphenyl. *J Nutr* 129: 1060–1067, 1999.
25. De Bont R and van Larebeke N. Endogenous DNA damage in humans: a review of quantitative data. *Mutagenesis* 19: 169–185, 2004.
26. Dohi Y, Ikura T, Hoshikawa Y, Katoh Y, Ota K, Nakanome A, Muto A, Omura S, Ohta T, Ito A, Yoshida M, Noda T, and Igarashi K. Bach1 inhibits oxidative stress-induced cellular senescence by impeding p53 function on chromatin. *Nat Struct Mol Biol* 15: 1246–1254, 2008.
27. Donald SP, Sun XY, Hu CA, Yu J, Mei JM, Valle D, and Phang JM. Proline oxidase, encoded by p53-induced gene-6, catalyzes the generation of proline-dependent reactive oxygen species. *Cancer Res* 61: 1810–1815, 2001.
28. Drane P, Bravard A, Bouvard V, and May E. Reciprocal down-regulation of p53 and SOD2 gene expression-implication in p53 mediated apoptosis. *Oncogene* 20: 430–439, 2001.
29. Droge W. Oxidative stress and ageing: is ageing a cysteine deficiency syndrome? *Phil Trans R Soc Lond B Biol Sci* 360: 2355–2372, 2005.
30. el-Deiry WS, Harper JW, O'Connor PM, Velculescu VE, Canman CE, Jackman J, Pietenpol JA, Burrell M, Hill DE, and Wang Y, et al. WAF1/CIP1 is induced in p53-mediated G1 arrest and apoptosis. *Cancer Res* 54: 1169–1174, 1994.
31. el-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, Lin D, Mercer WE, Kinzler KW, and Vogelstein B. WAF1, a potential mediator of p53 tumor suppression. *Cell* 75: 817–825, 1993.
32. Ethen CM, Reilly C, Feng X, Olsen TW, and Ferrington DA. Age-related macular degeneration and retinal protein modification by 4-hydroxy-2-nonenal. *Invest Ophthalmol Vis Sci* 48: 3469–3479, 2007.
33. Fan C, He L, Kapoor A, Gillis A, Rybak AP, Cutz JC, and Tang D. Bmi1 promotes prostate tumorigenesis via inhibiting p16(INK4A) and p14(ARF) expression. *Biochim Biophys Acta* 1782: 642–648, 2008.
34. Faraonio R, Vergara P, Di Marzo D, Pierantoni MG, Napolitano M, Russo T, and Cimino F. p53 suppresses the Nrf2-dependent transcription of antioxidant response genes. *J Biol Chem* 281: 39776–39784, 2006.
35. Freeman J, Schmidt S, Scharer E, and Iggo R. Mutation of conserved domain II alters the sequence specificity of DNA binding by the p53 protein. *EMBO J* 13: 5393–5400, 1994.
36. Galbiati F, Volonte D, Liu J, Capozza F, Frank PG, Zhu L, Pestell RG, Lisanti MP. Caveolin-1 expression negatively regulates cell cycle progression by inducing G(0)/G(1) arrest via a p53/p21(WAF1/Cip1)-dependent mechanism. *Mol Biol Cell* 12: 2229–2244, 2001.
37. Garinis GA, van der Horst GTJ, Vijg JHJ, and Hoeijmakers J. DNA damage and ageing: new-age ideas for an age-old problem. *Nat Cell Biol* 10: 1241–1247, 2008.
38. Gaziano JM. Vitamin E and cardiovascular disease: observational studies. *Ann N Y Acad Sci* 1031: 280–291, 2004.
39. Gems D and Partridge L. Insulin/IGF signalling and ageing: seeing the bigger picture. *Curr Opin Genet Dev* 11: 287–292, 2001.

40. Gentry A and Venkatachalam S. Complicating the role of p53 in aging. *Aging Cell* 4: 157–160, 2005.
41. Giannakou ME and Partridge L. The interaction between FOXO and SIRT1: tipping the balance towards survival. *Trends Cell Biol* 14: 408–412, 2004.
42. Guarente L. Sir2 links chromatin silencing, metabolism, and aging. *Genes Dev* 14: 1021–1026, 2000.
43. Gudkov A. Microarray analysis of p53-mediated transcription: multi-thousand piece puzzle or invitation to collective thinking. *Cancer Biol Ther* 2: 444–445, 2003.
44. Gupta A, Hasan M, Chander R, and Kapoor NK. Age-related elevation of lipid peroxidation products: diminution of superoxide dismutase activity in the central nervous system of rats. *Gerontology* 37: 305–309, 1991.
45. Hamilton ML, Van Remmen H, Drake JA, Yang H, Guo ZM, Kewitt K, Walter CA, and Richardson A. Does oxidative damage to DNA increase with age? *Proc Natl Acad Sci U S A* 98: 10469–10474, 2001.
46. Harman D. Aging: a theory based on free radical and radiation chemistry. *J Gerontol* 11: 298–300, 1956.
47. Haupt Y, Maya R, Kazaz A, and Oren M. Mdm2 promotes the rapid degradation of p53. *Nature* 387: 296–299, 1997.
48. Heo JY, Jing K, Song KS, Seo KS, Park JH, Kim JS, Jung YJ, Hur GM, Jo DY, Kweon GR, Yoon WH, Lim K, Hwang BD, Jeon BH, and Park JI. Downregulation of APE1/Ref-1 is involved in the senescence of mesenchymal stem cells. *Stem Cells* 27: 1455–1462, 2009.
49. Hercberg S, Galan P, Preziosi P, Bertrais S, Mennen L, Malvy D, Roussel AM, Favier A, and Briancon S. The SU.VI.MAX Study: a randomized, placebo-controlled trial of the health effects of antioxidant vitamins and minerals. *Arch Intern Med* 164: 2335–2342, 2004.
50. Hermeking H, Lengauer C, Polyak K, He TC, Zhang L, Thiagalingam S, Kinzler KW, and Vogelstein B. 14-3-3 sigma is a p53-regulated inhibitor of G2/M progression. *Mol Cell* 1: 3–11, 1997.
51. Honda R, Tanaka H, and Yasuda H. Oncoprotein MDM2 is a ubiquitin ligase E3 for tumor suppressor p53. *FEBS Lett* 420: 25–27, 1997.
52. Honda Y and Honda S. Oxidative stress and life span determination in the nematode *Caenorhabditis elegans*. *Ann N Y Acad Sci* 959: 466–474, 2002.
53. Huang J, Logsdon N, Schmieg FI, and Simmons DT. p53-mediated transcription induces resistance of DNA to UV inactivation. *Oncogene* 17: 401–411, 1998.
54. Hussain SP, Amstad P, He P, Robles A, Lupold S, Kaneko I, Ichimiya M, Sengupta S, Mechanic L, Okamura S, Hofseth LJ, Moake M, Nagashima M, Forrester KS, and Harris CC. p53-induced up-regulation of MnSOD and GPx but not catalase increases oxidative stress and apoptosis. *Cancer Res* 64: 2350–2356, 2004.
55. Jeffers JR, Parganas E, Lee Y, Yang C, Wang J, Brennan J, MacLean KH, Han J, Chittenden T, Ihle JN, McKinnon PJ, Cleveland JL, and Zambetti GP. Puma is an essential mediator of p53-dependent and -independent apoptotic pathways. *Cancer Cell* 4: 321–328, 2003.
56. Katic M and Kahn CR. The role of insulin and IGF-1 signaling in longevity. *Cell Mol Life Sci* 62: 320–343, 2005.
57. Kim J, Takahashi M, Shimizu T, Shirasawa T, Kajita M, Kanayama A, and Miyamoto Y. Effects of a potent antioxidant, platinum nanoparticle, on the lifespan of *Caenorhabditis elegans*. *Mech Ageing Dev* 129: 322–331, 2008.
58. Kondoh H, Lleonart ME, Gil J, Wang J, Degan P, Peters G, Martinez D, Carnero A, and Beach D. Glycolytic enzymes can modulate cellular life span. *Cancer Res* 65: 177–185, 2005.
59. Lamb DJ, Tickner ML, Hourani SM, and Ferns GA. Dietary copper supplements modulate aortic superoxide dismutase, nitric oxide and atherosclerosis. *Int J Exp Pathol* 86: 247–255, 2005.
60. Langley E, Pearson M, Faretta M, Bauer UM, Frye RA, Minucci S, Pelicci PG, and Kouzarides T. Human SIR2 deacetylates p53 and antagonizes PML/p53-induced cellular senescence. *EMBO J* 21: 2383–2396, 2002.
61. Lemon JA, Rollo CD, and Boreham DR. Elevated DNA damage in a mouse model of oxidative stress: impacts of ionizing radiation and a protective dietary supplement. *Mutagenesis* 23: 473–482, 2008.
62. Lenaz G. Role of mitochondria in oxidative stress and ageing. *Biochim Biophys Acta* 1366: 53–67, 1998.
63. Lenaz G, Bovina C, Formigini G, and Parenti Castelli G. Mitochondria, oxidative stress, and antioxidant defences. *Acta Biochim Pol* 46: 1–21, 1999.
64. Liu DP, Ou L, Clemenson Jr GD, Chao C, Lutske ME, Zambetti GP, Gage FH, and Xu Y. Puma is required for p53-induced depletion of adult stem cells. *Nat Cell Biol* 12: 993–998, 2010.
65. Linnane AW, Marzuki S, Ozawa T, and Tanaka M. Mitochondrial DNA mutations as an important contributor to ageing and degenerative diseases. *Lancet* 1: 642–645, 1989.
66. Liu Z, Lu H, Shi H, Du Y, Yu J, Gu S, Chen X, Liu KJ, and Hu CA. PUMA overexpression induces reactive oxygen species generation and proteasome-mediated stathmin degradation in colorectal cancer cells. *Cancer Res* 65: 1647–1654, 2005.
67. Lopez-Torres M and Barja G. Calorie restriction, oxidative stress and longevity. *Rev Esp Geriatr Gerontol* 43: 252–260, 2008.
68. Lu H and Levine AJ. Human TAFII31 protein is a transcriptional coactivator of the p53 protein. *Proc Natl Acad Sci U S A* 92: 5154–5158, 1995.
69. Lu H, Lin J, Chen J, and Levine AJ. The regulation of p53-mediated transcription and the roles of hTAFII31 and mdm-2. *Harvey Lect* 90: 81–93, 1994.
70. Machado A, Ayala A, Gordillo E, Revilla E, and Santa Maria C. Relationship between enzymatic activity loss and post-translational protein modification in aging. *Arch Gerontol Geriatr* 12: 187–197, 1991.
71. Macip S, Igarashi M, Berggren P, Yu J, Lee SW, and Aaronson SA. Influence of induced reactive oxygen species in p53-mediated cell fate decisions. *Mol Cell Biol* 23: 8576–8585, 2003.
72. Maier B, Gluba W, Bernier B, Turner T, Mohammad K, Guise T, Sutherland A, Thorner M, and Scrabble H. Modulation of mammalian life span by the short isoform of p53. *Genes Dev* 18: 306–319, 2004.
73. Mansur CP. The regulation and function of the p53 tumor suppressor. *Adv Dermatol* 13: 121–166, 1997.
74. Matheu A, Maraver A, Klatt P, Flores I, Garcia-Cao I, Borrás C, Flores JM, Vina J, Blasco MA, and Serrano M. Delayed ageing through damage protection by the Arf/p53 pathway. *Nature* 448: 375–379, 2007.
75. Mattson MP, Maudsley S, and Martin B. A neural signaling triumvirate that influences ageing and age-related disease: insulin/IGF-1, BDNF and serotonin. *Ageing Res Rev* 3: 445–464, 2004.



76. Meissner C. Mutations of mitochondrial DNA: cause or consequence of the ageing process? *Z Gerontol Geriatr* 40: 325–333, 2007.
77. Melov S, Schneider JA, Day BJ, Hinerfeld D, Coskun P, Mirra SS, Crapo JD, and Wallace DC. A novel neurological phenotype in mice lacking mitochondrial manganese superoxide dismutase. *Nat Genet* 18: 159–163, 1998.
78. Mendrysa SM, O'Leary KA, McElwee MK, Michalowski J, Eisenman RN, Powell DA, and Perry ME. Tumor suppression and normal aging in mice with constitutively high p53 activity. *Genes Dev* 20: 16–21, 2006.
79. Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, Lanfranccone L, and Pelicci PG. The p66shc adaptor protein controls oxidative stress response and life span in mammals. *Nature* 402: 309–313, 1999.
80. Migliaccio E, Mele S, Salcini AE, Pelicci G, Lai KM, Superti-Furga G, Pawson T, Di Fiore PP, Lanfranccone L, and Pelicci PG. Opposite effects of the p52shc/p46shc and p66shc splicing isoforms on the EGF receptor-MAP kinase-fos signalling pathway. *EMBO J* 16: 706–716, 1997.
81. Mocchegiani E, Malavolta M, Muti E, Costarelli L, Cipriano C, Piacenza F, Tesei S, Giacconi R, and Lattanzio F. Zinc, metallothioneins and longevity: interrelationships with niacin and selenium. *Curr Pharm Des* 14: 2719–2732, 2008.
82. Montaner B, O'Donovan P, Reelfs O, Perrett CM, Zhang X, Xu YZ, Ren X, Macpherson P, Frith D, and Karran P. Reactive oxygen-mediated damage to a human DNA replication and repair protein. *EMBO Rep* 8: 1074–1079, 2007.
83. Montine TJ, Neely MD, Quinn JF, Beal MF, Markesbery WR, Roberts LJ, and Morrow JD. Lipid peroxidation in aging brain and Alzheimer's disease. *Free Radic Biol Med* 33: 620–626, 2002.
84. Nakamizo A, Amano T, Zhang W, Zhang XQ, Ramdas L, Liu TJ, Bekele BN, Shono T, Sasaki T, Benedict WF, Sawaya R, and Lang FF. Phosphorylation of Thr18 and Ser20 of p53 in Ad-p53-induced apoptosis. *Neurol Oncol* 10: 275–291, 2008.
85. Neilsen PM, Cheney KM, Li CW, Chen JD, Cawrse JE, Schulz RB, Powell JA, Kumar R, and Callen DF. Identification of ANKRD11 as a p53 coactivator. *J Cell Sci* 121: 3541–3552, 2008.
86. Neumann CA, Krause DS, Carman CV, Das S, Dubey DP, Abraham JL, Bronson RT, Fujiwara Y, Orkin SH, and Van Etten RA. Essential role for the peroxiredoxin Prdx1 in erythrocyte antioxidant defence and tumour suppression. *Nature* 424: 561–565, 2003.
87. Nowak M, Swietochowska E, Wielkoszynski T, Marek B, Karpe J, Gorski J, Glogowska-Szelag J, Kos-Kudla B, and Ostrowska Z. Changes in blood antioxidants and several lipid peroxidation products in women with age-related macular degeneration. *Eur J Ophthalmol* 13: 281–286, 2003.
88. Ogata M. Acatalasemia. *Hum Genet* 86: 331–340, 1991.
89. Oguro H, Iwama A, Morita Y, Kamijo T, van Lohuizen M, and Nakauchi H. Differential impact of Ink4a and Arf on hematopoietic stem cells and their bone marrow microenvironment in Bmi1-deficient mice. *J Exp Med* 203: 2247–2253, 2006.
90. Orsini F, Migliaccio E, Moroni M, Contursi C, Raker VA, Piccini D, Martin-Padura I, Pelliccia G, Trinei M, Bono M, Puri C, Tacchetti C, Ferrini M, Mannucci R, Nicoletti I, Lanfranccone L, Giorgio M, and Pelicci PG. The life span determinant p66Shc localizes to mitochondria where it associates with mitochondrial heat shock protein 70 and regulates trans-membrane potential. *J Biol Chem* 279: 25689–25695, 2004.
91. Pani G, Bedogni B, Anzevino R, Colavitti R, Palazzotti B, Borrello S, and Galeotti T. Deregulated manganese superoxide dismutase expression and resistance to oxidative injury in p53-deficient cells. *Cancer Res* 60: 4654–4660, 2000.
92. Pavletich NP, Chambers KA, and Pabo CO. The DNA-binding domain of p53 contains the four conserved regions and the major mutation hot spots. *Genes Dev* 7: 2556–2564, 1993.
93. Pelicci G, Lanfranccone L, Grignani F, McGlade J, Cavallo F, Forni G, Nicoletti I, Grignani F, Pawson T, and Pelicci PG. A novel transforming protein (SHC) with an SH2 domain is implicated in mitogenic signal transduction. *Cell* 70: 93–104, 1992.
94. Pinkston JM, Garigan D, Hansen M, and Kenyon C. Mutations that increase the life span of *C. elegans* inhibit tumor growth. *Science* 313: 971–975, 2006.
95. Polyak K, Xia Y, Zweier JL, Kinzler KW, and Vogelstein B. A model for p53-induced apoptosis. *Nature* 389: 300–305, 1997.
96. Poulsen HE. Oxidative DNA modifications. *Exp Toxicol Pathol* 57 suppl 1: 161–169, 2005.
97. Pratico D. Lipid peroxidation and the aging process. *Sci Aging Knowledge Environ* 50: 1–4, 2002.
98. Reaume AG, Elliott JL, Hoffman EK, Kowall NW, Ferrante RJ, Siwek DF, Wilcox HM, Flood DG, Beal MF, Brown RH Jr, Scott RW, and Snider WD. Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nat Genet* 13: 43–47, 1996.
99. Rhee SG, Chae HZ, and Kim K. Peroxiredoxins: a historical overview and speculative preview of novel mechanisms and emerging concepts in cell signaling. *Free Radic Biol Med* 38: 1543–1552, 2005.
100. Sablina AA, Budanov AV, Ilyinskaya GV, Agapova LS, Kravchenko JE, and Chumakov PM. The antioxidant function of the p53 tumor suppressor. *Nat Med* 11: 1306–1313, 2005.
101. Sakaguchi K, Herrera JE, Saito S, Miki T, Bustin M, Vassilev A, Anderson CW, and Appella E. DNA damage activates p53 through a phosphorylation-acetylation cascade. *Genes Dev* 12: 2831–2841, 1998.
102. Sandor J, Ambrus T, and Ember I. The function of the p53 gene suppressor in carcinogenesis. *Orv Hetil* 136: 1875–1883, 1995.
103. Schoneich C. Protein modification in aging: an update. *Exp Gerontol* 41: 807–812, 2006.
104. Scolnick DM, Chehab NH, Stavridi ES, Lien MC, Caruso L, Moran E, Berger SL, and Halazonetis TD. CREB-binding protein and p300/CBP-associated factor are transcriptional coactivators of the p53 tumor suppressor protein. *Cancer Res* 57: 3693–3696, 1997.
105. Sedding DG. FoxO transcription factors in oxidative stress response and ageing: a new fork on the way to longevity? *Biol Chem* 389: 279–283, 2008.
106. Selenius M, Rundlof AK, Olm E, Fernandes AP, and Bjornstedt M. Selenium and the selenoprotein thioredoxin reductase in the prevention, treatment and



- diagnostics of cancer. *Antioxid Redox Signal* 12: 867–880, 2009.
107. Sesso HD, Buring JE, Christen WG, Kurth T, Belanger C, MacFadyen J, Bubes V, Manson JE, Glynn RJ, and Gaziano JM. Vitamins E and C in the prevention of cardiovascular disease in men: the Physicians' Health Study II randomized controlled trial. *JAMA* 300: 2123–2133, 2008.
  108. Shinohara R, Mano T, Nagasaka A, Hayashi R, Uchimura K, Nakano I, Watanabe F, Tsugawa T, Makino M, Kakiyama H, Nagata M, Iwase K, Ishizuki Y, and Itoh M. Lipid peroxidation levels in rat cardiac muscle are affected by age and thyroid status. *J Endocrinol* 164: 97–102, 2000.
  109. Siedlak SL, Casadesus G, Webber KM, Pappolla MA, Atwood CS, Smith MA, and Perry G. Chronic antioxidant therapy reduces oxidative stress in a mouse model of Alzheimer's disease. *Free Radic Res* 43: 156–164, 2009.
  110. Sonntag WE, Lynch C, Thornton P, Khan A, Bennett S, and Ingram R. The effects of growth hormone and IGF-1 deficiency on cerebrovascular and brain ageing. *J Anat* 197: 575–585, 2000.
  111. Soskic V, Groebe K, and Schrattenholz A. Nonenzymatic posttranslational protein modifications in ageing. *Exp Gerontol* 43: 247–257, 2008.
  112. Stadtman ER. Protein modification in aging. *J Gerontol* 43: B112–B120, 1988.
  113. Stewart RR and Bewley JD. Lipid peroxidation associated with accelerated aging of soybean axes. *Plant Physiol* 65: 245–248, 1980.
  114. Takubo K, Ohmura M, Azuma M, Nagamatsu G, Yamada W, Arai F, Hirao A, and Suda T. Stem cell defects in ATM-deficient undifferentiated spermatogonia through DNA damage-induced cell-cycle arrest. *Cell Stem Cell* 2: 170–182, 2008.
  115. Tan M, Li S, Swaroop M, Guan K, Oberley LW, and Sun Y. Transcriptional activation of the human glutathione peroxidase promoter by p53. *J Biol Chem* 274: 12061–12066, 1999.
  116. Terzioglu M and Larsson NG. Mitochondrial dysfunction in mammalian ageing. *Novartis Found Symp* 287: 197–208, 2007.
  117. Toescu EC, Myronova N, and Verkhatsky A. Age-related structural and functional changes of brain mitochondria. *Cell Calcium* 28: 329–338, 2000.
  118. Trinei M, Giorgio M, Cicalese A, Barozzi S, Ventura A, Migliaccio E, Milia E, Padura IM, Raker VA, Maccarana M, Petronilli V, Minucci S, Bernardi P, Lanfranccone L, and Pelicci PG. A p53-p66Shc signalling pathway controls intracellular redox status, levels of oxidation-damaged DNA and oxidative stress-induced apoptosis. *Oncogene* 21: 3872–3878, 2002.
  119. Tyner SD, Venkatachalam S, Choi J, Jones S, Ghebranious N, Igelmann H, Lu X, Soron G, Cooper B, Brayton C, Hee Park S, Thompson T, Karsenty G, Bradley A, and Donehower LA. p53 mutant mice that display early ageing-associated phenotypes. *Nature* 415: 45–53, 2002.
  120. van Heemst D, Mooijaart SP, Beekman M, Schreuder J, de Craen AJ, Brandt BW, Slagboom PE, and Westendorp RG. Variation in the human TP53 gene affects old age survival and cancer mortality. *Exp Gerontol* 40: 11–15, 2005.
  121. Vanfleteren JR and Braeckman BP. Mechanisms of life span determination in *Caenorhabditis elegans*. *Neurobiol Aging* 20: 487–502, 1999.
  122. Vaziri H, Dessain SK, Ng Eaton E, Imai SI, Frye RA, Pandita TK, Guarente L, and Weinberg RA. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell* 107: 149–159, 2001.
  123. Videla LA, Fernandez V, and Valenzuela A. Age-dependent changes in rat liver lipid peroxidation and glutathione content induced by acute ethanol ingestion. *Cell Biochem Funct* 5: 273–280, 1987.
  124. Viner RI, Ferrington DA, Williams TD, Bigelow DJ, and Schoneich C. Protein modification during biological aging: selective tyrosine nitration of the SERCA2a isoform of the sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase in skeletal muscle. *Biochem J* 340: 657–669, 1999.
  125. Vivekananthan DP, Penn MS, Sapp SK, Hsu A, and Topol EJ. Use of antioxidant vitamins for the prevention of cardiovascular disease: meta-analysis of randomised trials. *Lancet* 361: 2017–2023, 2003.
  126. Volonte D, Zhang K, Lisanti MP, and Galbiati F. Expression of caveolin-1 induces premature cellular senescence in primary cultures of murine fibroblasts. *Mol Biol Cell* 13: 2502–2517, 2002.
  127. Walker DR, Bond JP, Tarone RE, Harris CC, Makalowski W, Boguski MS, and Greenblatt MS. Evolutionary conservation and somatic mutation hotspot maps of p53: correlation with p53 protein structural and functional features. *Oncogene* 18: 211–218, 1999.
  128. Wood ZA, Schroder E, Robin Harris J, and Poole LB. Structure, mechanism and regulation of peroxiredoxins. *Trends Biochem Sci* 28: 32–40, 2003.
  129. Xu Y. Regulation of p53 responses by post-translational modifications. *Cell Death Differ* 10: 400–403, 2003.
  130. Yoon KA, Nakamura Y, and Arakawa H. Identification of ALDH4 as a p53-inducible gene and its protective role in cellular stresses. *J Hum Genet* 49: 134–140, 2004.
  131. Zaky A, Busso C, Izumi T, Chattopadhyay R, Bassiouny A, Mitra S, and Bhakat KK. Regulation of the human AP-endonuclease (APE1/Ref-1) expression by the tumor suppressor p53 in response to DNA damage. *Nucleic Acids Res* 36: 1555–1566, 2008.
  132. Zamocky M and Koller F. Understanding the structure and function of catalases: clues from molecular evolution and in vitro mutagenesis. *Prog Biophys Mol Biol* 72: 19–66, 1999.
  133. Zhu D, Wu J, Spee C, Ryan SJ, and Hinton DR. BMP4 mediates oxidative stress-induced retinal pigment epithelial cell senescence and is overexpressed in age-related macular degeneration. *J Biol Chem* 284: 9529–9539, 2009.

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**Abbreviations Used**

ALDH = aldehyde dehydrogenase  
APE/Ref1 = apurinic/aprimidinic endonuclease/  
redox factor-1  
BAI1 = brain-specific angiogenesis inhibitor 1  
Dmp53 = *Drosophila melanogaster* p53  
ES cell = embryonic stem cell  
FoxO = forkhead box O  
GPX = glutathione peroxidase  
GST = glutathione S-transferase  
hBMSCs = human bone marrow-derived  
mesenchymal stem cells  
MEF = mouse embryonic fibroblast

NQO1 = NAD(P)H dehydrogenase [quinone] 1  
Nrf1 = NF-E2-related factor 2  
PGM = phosphoglycerate mutase  
PIG3 = p53-inducible gene 3  
Puma = p53 upregulated modulator of apoptosis  
ROS = reactive oxygen species  
SGK = serum- and glucocorticoid-inducible  
kinase  
SOD = superoxide dismutase  
TIGAR = TP53-induced glycolysis and apoptosis  
regulator  
TP53INP1 = tumor protein 53-induced nuclear  
protein 1  
TSP1 = thrombospondin-1

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1. James R. Van Brocklyn, Joseph B. Williams. 2012. The control of the balance between ceramide and sphingosine-1-phosphate by sphingosine kinase: Oxidative stress and the seesaw of cell survival and death. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* **163**:1, 26-36. [[CrossRef](#)]
2. Mari Sawamoto, Takafumi Imai, Mana Umeda, Koji Fukuda, Takao Kataoka, Shigeru Taketani. 2012. The p53-Dependent Expression of Frataxin Controls 5-Aminolevulinic Acid-Induced Accumulation of Protoporphyrin IX and Photo-Damage in Cancerous Cells. *Photochemistry and Photobiology* n/a-n/a. [[CrossRef](#)]
3. Clement T.Y. Chan, Yan Ling Joy Pang, Wenjun Deng, I. Ramesh Babu, Madhu Dyavaiah, Thomas J. Begley, Peter C. Dedon. 2012. Reprogramming of tRNA modifications controls the oxidative stress response by codon-biased translation of proteins. *Nature Communications* **3**, 937. [[CrossRef](#)]
4. Su-Jeong Kim, Yun-Jong Park, Young J. Oh. 2012. Proteomic analysis reveals a protective role for DJ-1 during 6-hydroxydopamine-induced cell death. *Biochemical and Biophysical Research Communications* **422**:1, 8-14. [[CrossRef](#)]
5. Xuemei Fu, Yang Xu. 2012. Challenges to the clinical application of pluripotent stem cells: towards genomic and functional stability. *Genome Medicine* **4**:6, 55. [[CrossRef](#)]
6. Vasileios Kordinas, Chrysoula Nicolaou, George Tsirpanlis, Anastasios Ioannidis, Sotiris Bersimis, Nikos Sabanis, Eleni Fragou, Konstantina Tsiolaki, Stylianos Chatzipanagiotou. 2012. Transcription of the Tumor Suppressor Genes p53 and RB in Lymphocytes from Patients with Chronic Kidney Disease: Evidence of Molecular Senescence?. *International Journal of Nephrology* **2012**, 1-7. [[CrossRef](#)]
7. Antero Salminen, Kai Kaarniranta. 2011. AMP-activated protein kinase (AMPK) controls the aging process via an integrated signaling network. *Ageing Research Reviews* . [[CrossRef](#)]
8. Weixi Kong, Robert K. Kuester, Alfred Gallegos, I. Glenn Sipes. 2011. Induction of DNA damage in human urothelial cells by the brominated flame retardant 2,2-bis(bromomethyl)-1,3-propanediol: Role of oxidative stress. *Toxicology* **290**:2-3, 271-277. [[CrossRef](#)]
9. Nilanjana Maulik , Juan A. Sanchez . 2011. Risk Factors in Heart Disease: Therapeutic Interventions. *Antioxidants & Redox Signaling* **15**:7, 1765-1767. [[Citation](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]